

AD _____

Award Number: W81XWH-04-1-0136

TITLE: Development of a Sustained Antiplatelet, Antimicrobial
Delivery System for KSL Localized in the Oral Cavity

PRINCIPAL INVESTIGATOR: Doctor Patrick P. Deluca

CONTRACTING ORGANIZATION: University of Kentucky Research
Foundation
Lexington, Kentucky 40506-0057

REPORT DATE: May 2004

TYPE OF REPORT: Final, Phase I

PREPARED FOR: U.S. Army Medical Research and Materiel Command
Fort Detrick, Maryland 21702-5012

DISTRIBUTION STATEMENT: Approved for Public Release;
Distribution Unlimited

The views, opinions and/or findings contained in this report are those of the author(s) and should not be construed as an official Department of the Army position, policy or decision unless so designated by other documentation.

20050105 007

REPORT DOCUMENTATION PAGE

Form Approved
OMB No. 074-0188

Public reporting burden for this collection of information is estimated to average 1 hour per response, including the time for reviewing instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing this collection of information. Send comments regarding this burden estimate or any other aspect of this collection of information, including suggestions for reducing this burden to Washington Headquarters Services, Directorate for Information Operations and Reports, 1215 Jefferson Davis Highway, Suite 1204, Arlington, VA 22202-4302, and to the Office of Management and Budget, Paperwork Reduction Project (0704-0188), Washington, DC 20503

1. AGENCY USE ONLY (Leave blank)		2. REPORT DATE May 2004	3. REPORT TYPE AND DATES COVERED Final, Phase I (1 Jan 04 - 30 Apr 04)	
4. TITLE AND SUBTITLE Development of a Sustained Antiplatelet, Antimicrobial Delivery System for KSL Localized in the Oral Cavity			5. FUNDING NUMBERS W81XWH-04-1-0136	
6. AUTHOR(S) Doctor Patrick P. Deluca				
7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES) University of Kentucky Research Foundation Lexington, Kentucky 40506-0057 E-Mail: ppdelul@pop.uky.edu			8. PERFORMING ORGANIZATION REPORT NUMBER	
9. SPONSORING / MONITORING AGENCY NAME(S) AND ADDRESS(ES) U.S. Army Medical Research and Materiel Command Fort Detrick, Maryland 21702-5012			10. SPONSORING / MONITORING AGENCY REPORT NUMBER	
11. SUPPLEMENTARY NOTES				
12a. DISTRIBUTION / AVAILABILITY STATEMENT Approved for Public Release; Distribution Unlimited				12b. DISTRIBUTION CODE
13. ABSTRACT (Maximum 200 Words) No abstract provided.				
14. SUBJECT TERMS No subject terms provided.				15. NUMBER OF PAGES 17
				16. PRICE CODE
17. SECURITY CLASSIFICATION OF REPORT Unclassified	18. SECURITY CLASSIFICATION OF THIS PAGE Unclassified	19. SECURITY CLASSIFICATION OF ABSTRACT Unclassified	20. LIMITATION OF ABSTRACT Unlimited	

NSN 7540-01-280-5500

Standard Form 298 (Rev. 2-89)
Prescribed by ANSI Std. Z39-18
298-102

Table of Contents

Cover.....	1
SF 298.....	2
Table of Contents.....	3
Introduction.....	4
Body.....	5
Results.....	7
Scheduled Studies.....	15
Conclusions.....	
References.....	
Appendices.....	

Development of a Sustained Antiplaque Antimicrobial Delivery System for KSL Localized in the Oral Cavity

This report summarizes the progress since the beginning of research in February. Meetings were held with Dr. Kai Leung and Army personnel in February at the University of Kentucky and the chewing machine was delivered. Preformulation studies commenced and the Analytical Methodology was developed for KSL.

Stability assessment in buffered solutions at pH 4.0, 7.4 and 9.0 reveals excellent stability at 37°C through 7 days at all pHs. The robustness is demonstrated even at 55°C for more than one day at pH 7.4. In artificial saliva, pH 5.7, there was no detectable loss at 37°C for 3 days. Studies are underway in whole mucosal saliva and simulated gastric and intestinal fluids.

Concurrent with the stability studies, since retention of the KSL in the oral cavity for a sufficient period of time will depend on incorporation in polymeric microspheres, retention of the microspheres in the gum base will be assessed. These studies will be done with blank microspheres in a gum base provided.

Incorporation of KSL into microspheres as well as determining the binding of KSL to blank microspheres will be performed. Following these studies, KSL containing microspheres will be incorporated into the gum base and release studies undertaken.

A summary of the results to date follows. In addition to the PI, personnel working on the grant during this period of time were:

Dr. Dong Hee Na
Paolo Blasi
Dr. Yilmaz Capan

DEVELOPMENT OF A SUSTAINED ANTIPLAQUE, ANTIMICROBIAL DELIVERY SYSTEM FOR KSL LOCALIZED IN THE ORAL CAVITY

May 3, 2004

Studies being carried out within the context of the project

Phase I : Preformulation Studies

A- Analytical Methods Development

a. Active Ingredients

Analytical procedures to assess drug content, stability and release have been developed.

Stock solution: KSL 3.0 mg/mL in D.W.

Test concentration: KSL 200 µg/mL in each solution

Analysis: Reversed-phase HPLC with C-18 column

HPLC conditions:

- Column: Prosphere C-18 (4.6 x 250 mm, Alltech, Deerfield, IL)
- Flow rate: 1.2 mL/min
- Injection volume: 40 µL
- Mobile Phase A: Water with 0.1 % TFA
- Mobile Phase B: Acetonitrile with 0.1 % TFA
- Gradient: 80% A & 20% B to 70% A & 30% B in 8 min.
- Detection: UV 215 nm

B- Stability Assessment

1. pH Effect

KSL was tested at three pH conditions as described below:

- pH 4.0, 20 mM sodium acetate buffer
- pH 7.4, 20 mM sodium phosphate buffer
- pH 9.0, 20 mM sodium borate buffer

2. Temperature Effect

The KSL may be exposed to higher temperatures during fabrication, actual use and storage. Therefore, the stability of the KSL was studied at 25, 37 and 55 °C.

In addition to the stability studies conducted at three different pH values and three different temperatures, stability studies in artificial saliva, simulated gastric fluid (USP), and simulated intestinal fluid (USP) at 37°C will be carried out.

3. Artificial saliva at 37°C

The Artificial saliva was used for the in vitro release study in an attempt to simulate the actual conditions of use. The ingredients of the artificial saliva follows:

Sodium chloride – 0.844 g

Potassium chloride – 1.200 g

Calcium chloride dihydrate – 0.193 g

Magnesium chloride hexahydrate – 0.111 g

Patassium phosphate dibasic – 0.342 g

Water to make to 1 L and pH adjustment with HCl to $\text{pH } 5.7 \pm 0.1$

Sampling:

Samples for studies of pH and temperature effects were taken on 0, 1st, 3rd, 7th, 14th, 21st, and 28th days (triplicate). The stability study in artificial saliva was performed for 3 hours.

RESULTS

A. Analytical Method Development

1. HPLC method of KSL

Specificity of HPLC method

Under the described HPLC conditions, the standard of KSL in D.W. was detected as a single peak at the retention time of 7.0 min (Figure 1a). The degradation compounds of KSL produced at each pH conditions (55°C) could be resolved by the HPLC method (Figure 1b-d).

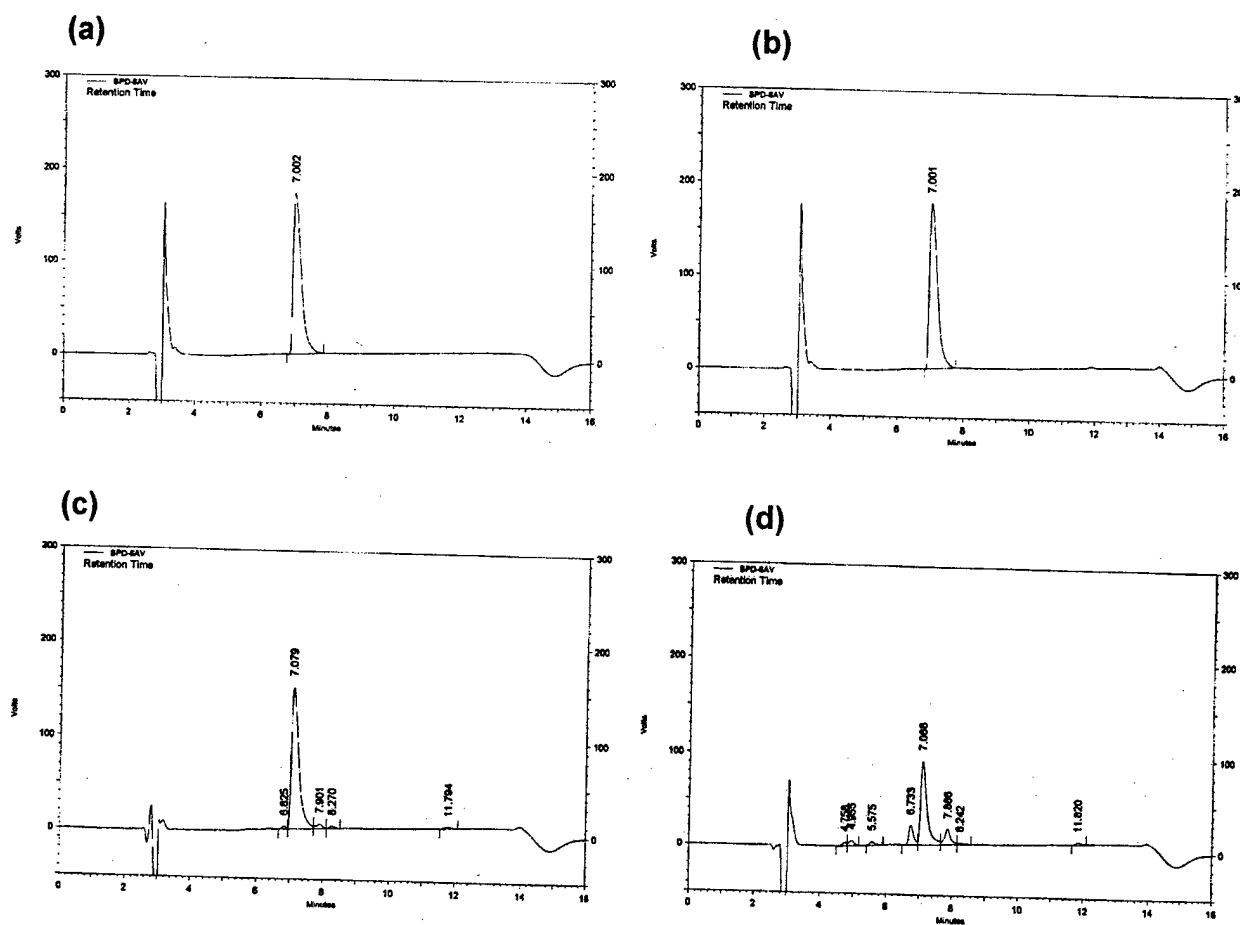


Figure 1. HPLC chromatograms of KSL in various aqueous solutions (a: KSL 200 µg/mL in D.W., b: KSL incubated at pH 4 (55°C) for 3 days, c: KSL incubated at pH 7.4 (55°C) for 3 days, d: KSL incubated at pH 9 (55°C) for 3 days).

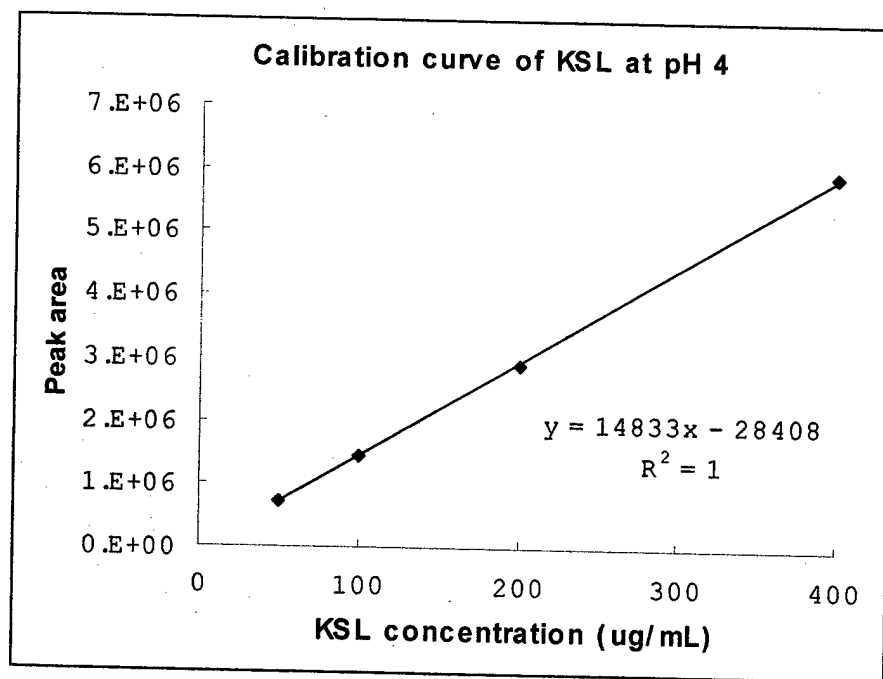
Calibration curve of KSL in each pH solutions

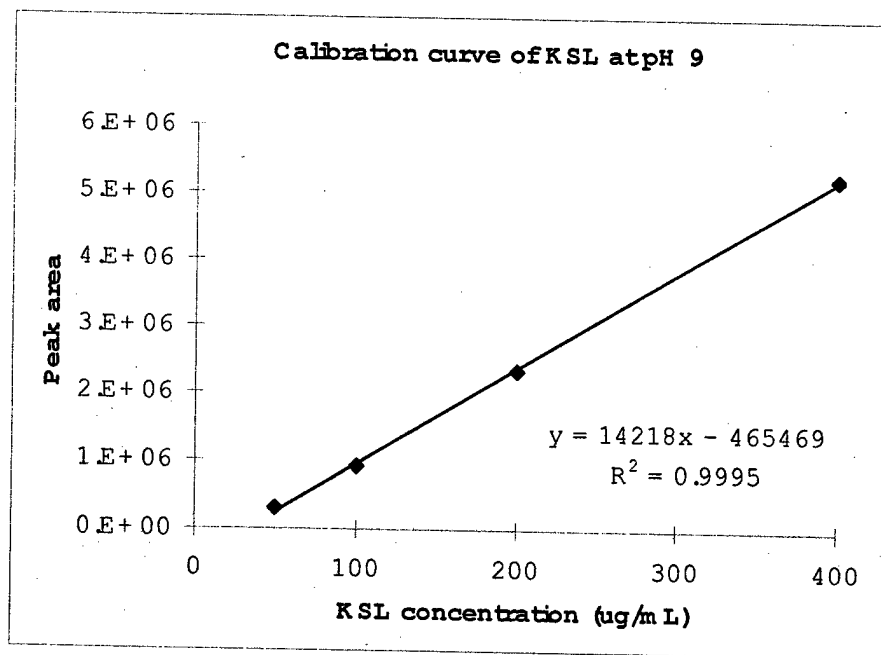
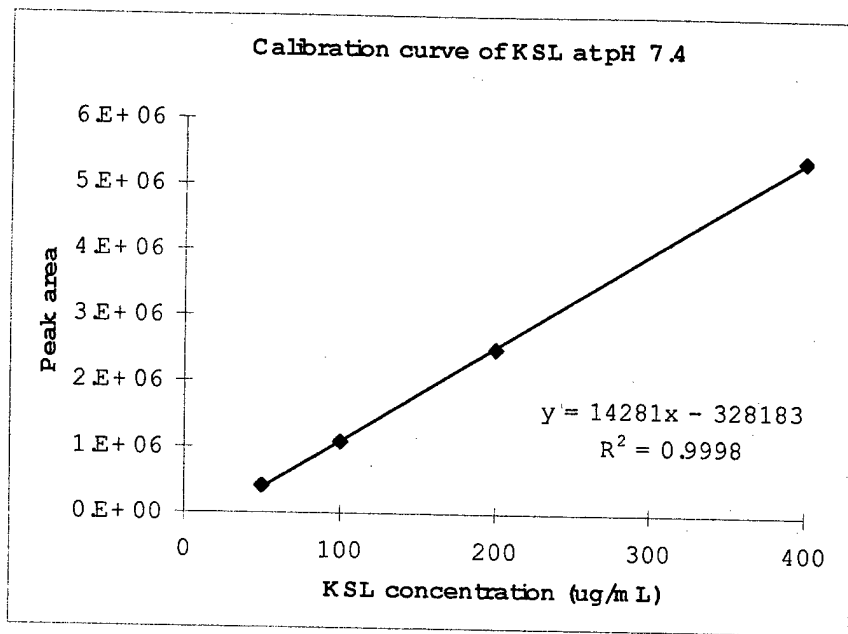
Table 1. Precision and linearity of KSL in various pH solutions

KSL concentration ($\mu\text{g/mL}$)	pH 4		pH 7.4		pH 9	
	Average peak area ¹	RSD (%) ²	Average peak area ¹	RSD (%) ²	Average peak area ¹	RSD (%) ²
50	725063	5.9	413753	10.1	301632	7.5
100	1455535	2.3	1089352	5.0	918281	5.3
200	2916429	0.7	2495015	1.8	2337003	3.3
400	5913898	2.5	5399814	1.6	5244962	2.8
Linearity	0.9999		0.9998		0.9995	

¹n=3 (inter-day)

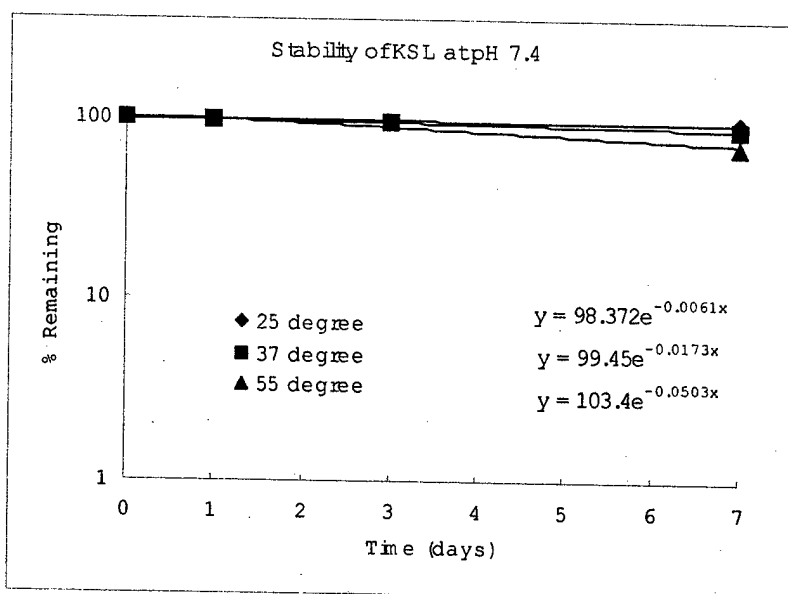
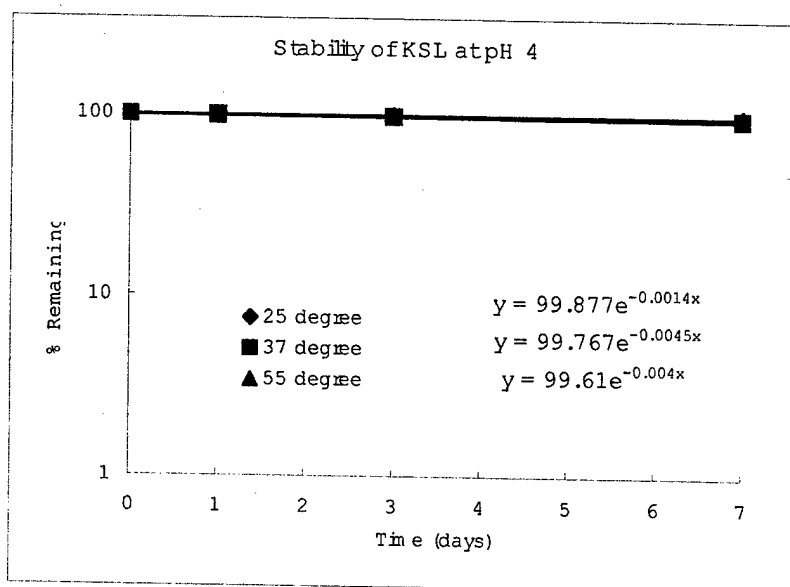
²Relative standard deviation





B. Stability of KSL in aqueous solutions

1. Degradation rate of KSL at various pH solutions



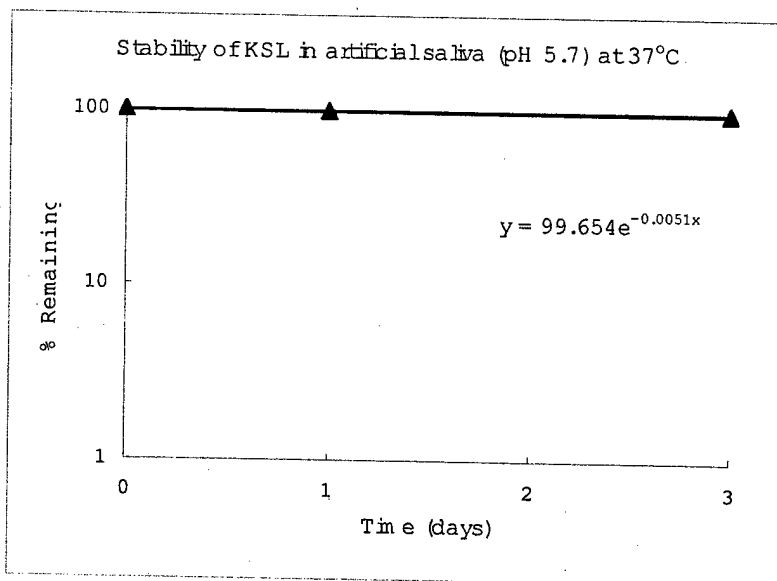
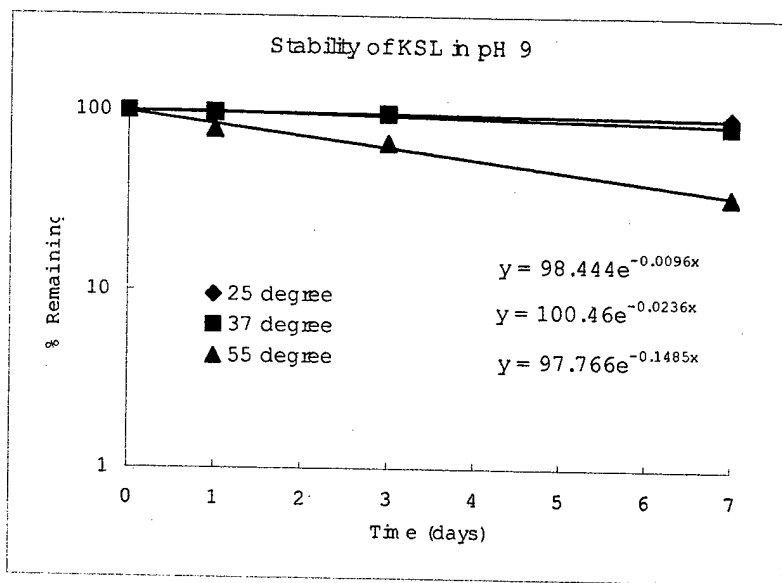
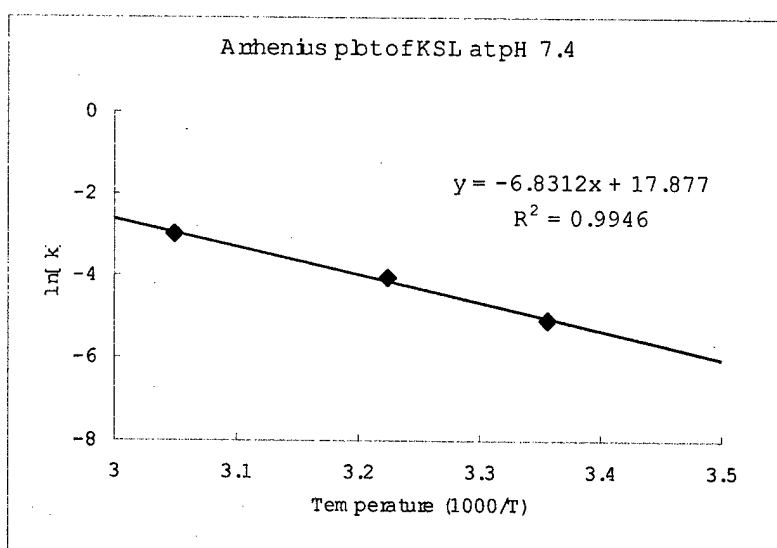
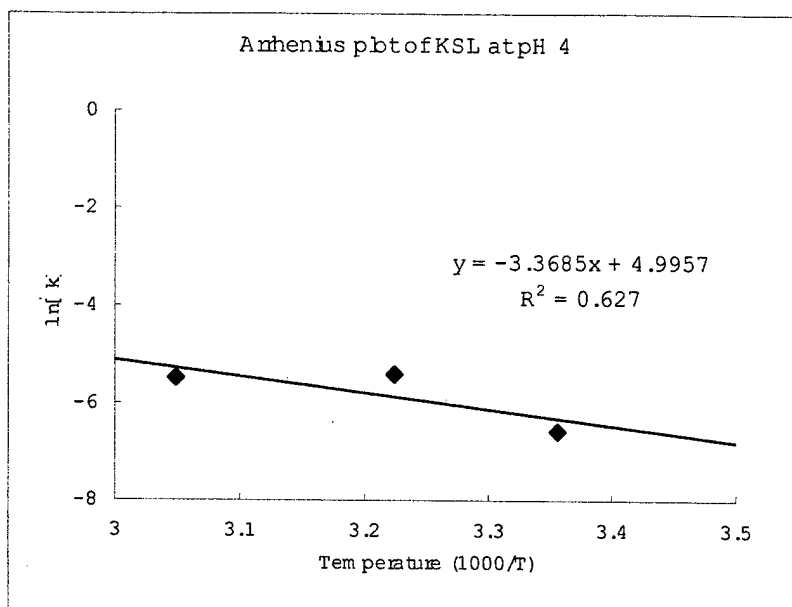


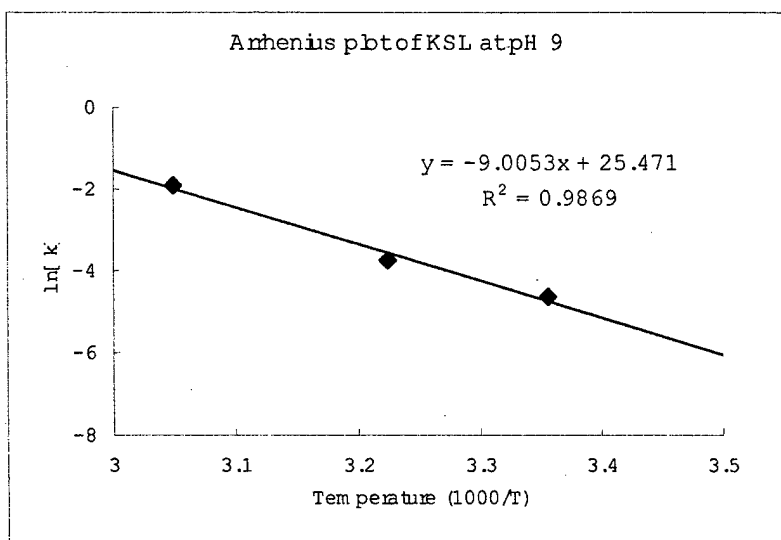
Table 1. Degradation constants of KSL

Temperature	pH 4	pH 7.4	pH 9	Artificial saliva
25°C	1.4×10^{-3}	6.1×10^{-3}	9.6×10^{-3}	-
37°C	4.5×10^{-3}	17.3×10^{-3}	23.6×10^{-3}	5.1×10^{-3}
55°C	4.2×10^{-3}	50.3×10^{-3}	148.5×10^{-3}	-
Ea (cal/degree/mole) ¹	6.693	13.574	17.894	-

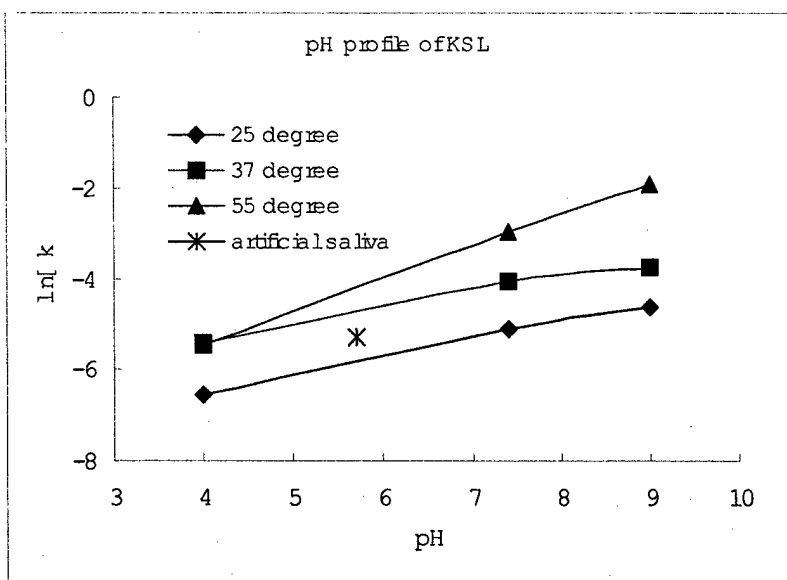
¹Activation energy

2. Arrhenius plot





3. pH effect on stability of KSL



SCHEDULED STUDIES

A. STABILITY STUDIES

1. Whole mucosal saliva. This has been provided by Dr. Leung
2. Simulated gastric and intestinal fluid

The formulations of the simulated gastric and intestinal fluids:

- *Simulated gastric fluid (USP)*

1 N HCl – 7 mL

NaCl – 2 g

Pepsin – 3.2 g

Water to make to 1 L, pH 1.2

- *Simulated intestinal fluid (USP) at 37°C*

Monobasic potassium phosphate – 6.8 g

Water – 500 mL

0.2 N NaOH – 190 mL

Pancreatin – 10 g

Water to make to 1 L, pH 6.8

Sampling: The stability study in simulated gastric fluid and intestinal fluid will be performed for 12 hours.

B. RETENTION OF MICROSPHERES

Incorporation of blank microspheres into Gum Base

On the basis of a 800 mg gum base, the following mixtures will be prepared.

10 % microspheres; 80 mg microspheres, 720 mg gum base

20 % microspheres; 160 mg microspheres, 640 mg gum base

40 % microspheres; 320 mg microspheres, 420 mg gum base

The mixtures will be subjected to the *chewing apparatus*

C. KSL ADSORPTION AND INCORPORATION INTO MICROSPHERES

C1-Adsorption Studies

Following the preparation of PLGA (Resomer 502 H) microspheres by a solvent extraction/ evaporation process, KSL will be adsorbed to the microsphere surface. Then, the amount of KSL adsorbed to the microsphere surface will be determined as *mg KSL/ mg microspheres*.

Maximum adsorption will be determined and release studies will be carried out in artificial saliva.

C2-Incorporation studies

KSL will be incorporated into microspheres during the fabrication of the microspheres at target drug loads of 10 and 20 % by a solvent extraction/evaporation process, and the actual drug content determined by HPLC. Release studies will be performed in artificial saliva.

D. PREPARATION OF KSL CHEWING GUM

The KSL-adsorbed and KSL-incorporated microspheres as well as KSL-containing chewing gum formulations will be prepared, and characterized for surface morphology by SEM and in vitro release in a buffer solution.

Calculations for determining KSL content:

Total weight of one chewing gum base is 800 mg.
If residence time for chewing gum is taken as 20 min,

Total saliva for 20 min: ~ 50 mL

ED₉₉ of KSL: 6.25 µg/mL (Ref. Concannon et al., J. Med. Microbiol., 2003, 52, 1083-1093)

6.25 µg/mL x 50 mL = 312.5 µg KSL required (minimum)

Calculation example:

If drug loading: x 10

312.5x10 = 3125 µg or 3.125 mg KSL

At a load of: 10% (drug/microsphere)

31.25 mg of microsphere (28.125 mg polymer + 3.125 mg KSL) will be added into 1 chewing gum.

**** We will start with a minimum of 31.25 mg /chewing gum and test up to a maximum amount that the chewing gum base can hold.**

The chewing gum formulations designed for testing

A. Conventional chewing gum formulations

Ingredient	A1	A2	A3
	Weight (%)	Weight (%)	Weight (%)
Gum base	52.6	29	30.5
Paraffin oil	3.4	-	-
Powdered sorbitol	40.8	43	-
Sorbitol solution 70%	-	21	-
Mannitol	0.9	-	-
Saccharin	0.1	-	-
Powdered sugar	-	-	50
Corn syrup	-	-	18
Aspartame	-	0.33	-
Glycerine	-	5	-
Lecithin	-	0.5	0.3
Flavor	2.1	1	1
Coloring	-	-	0.2
Sodium fluoride	0.1	-	-